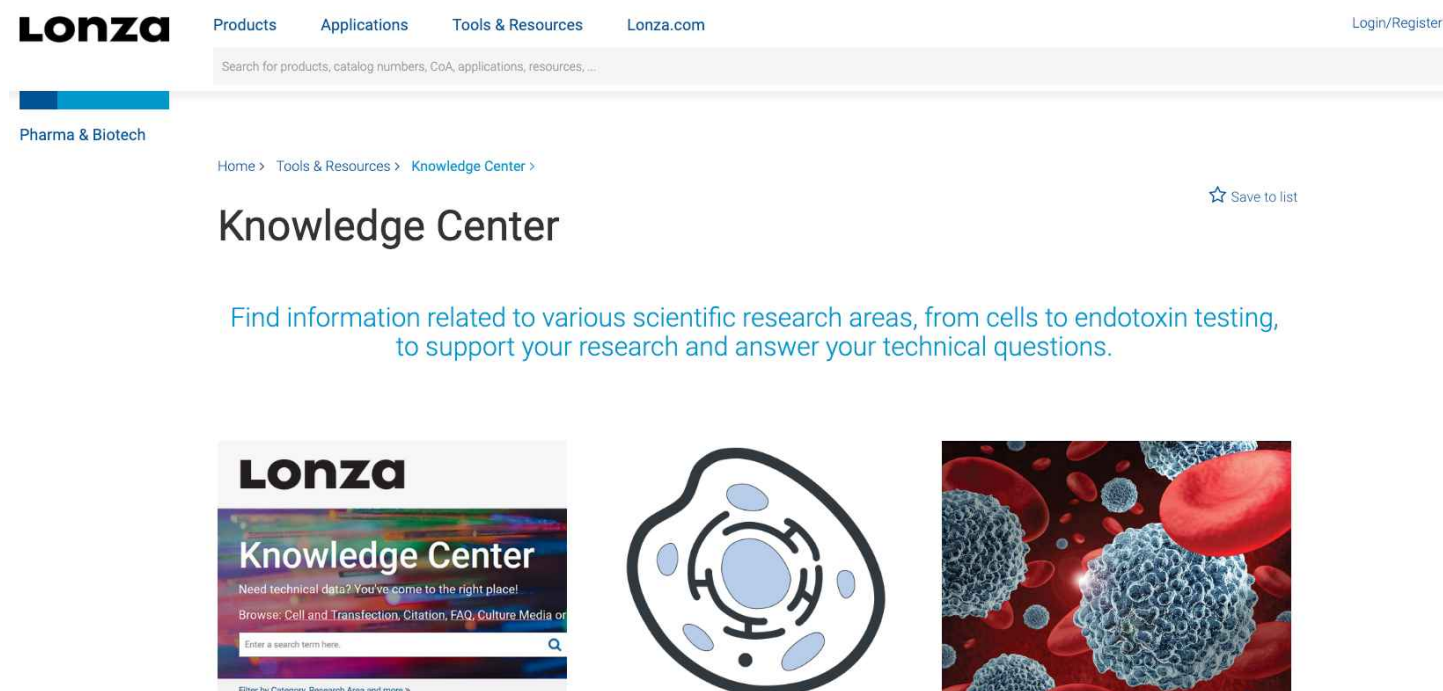


LONZA nucleofection information

If we need to perform nucleofection with our cells, we can search for more information from LONZA Knowledge Center:

https://bioscience.lonza.com/lonza_bs/CN/en/knowledge-center



And click the cell at the centre of above picture, then we will enter into the nucleofection knowledge list:

https://knowledge.lonza.com/search-results?search=*%&type=Cell%20Information&orderby=relevance

here you can choose what types of information you want to know →

here are more details for you to choose that you may want to know in your experiments →

here you can choose the research area you interested →

here are some information about the nucleofection system you want to use →

you can search key words for more information

if you find the cell line that you are interested, you can click it for more information

Cell Line	Description
NSC34	spinal cord neuroblastoma
Molt16	T cell lymphoma, Leukemic T cell line
IHH	immortalized hepatocyte
IM9	B-lymphoblastoid cells from the bone marrow of a woman with multiple myeloma but cells appear to be EBV+ B-lymphoblastoid cells' rather than myeloma-derived cells (see also Pellat-Deceunynk et al., Blood 86: 4001-4002, 1995)
mIMCD3	kidney, medulla; collecting duct; SV40 transformed
INS1 832/13	Insulin secreting cells (stably transfected with Plasmid coding human Proinsulin)
IOSE80	ovarian surface epithelium (OSE) cell line; SV40-immort. normal OSE cells

In this knowledge center, LONZA will give you some nucleofection experiment protocols for each cell line:

Lonza



Embryonic stem cell (ES), mouse

[Export](#)

Stem cells derived from mouse blastocyst.

Cell Type: Embryonic SC
Tissue Origin: embryo
Species: mouse
Research Area: Cancer Research/Cell Biology
Stem Cells
Cell Characteristics: Adherent

Transfection Information

[Lonza Optimized Protocol](#)

[Optimization Guideline](#)

Filter:

The table below shows data for the cell type and Nucleofector™ Platform selected. Those data are either based on Lonza Optimized Protocols or on results shared from customers who performed an optimization based on our guidelines. In case no data are shown for the selected Nucleofector™ Platform, please take a look at our [optimization strategy](#) to get further guidance on how to easily determine optimal Nucleofection conditions yourself.

Protocol	Kit	Program	Cells	Efficiency	Viable Cells	Substrate	Format	Platform
	P3	96-CG-104	5e4	85-95%	58-88%	Plasmid (general)	0.4 µg 20 µl	96-well
	ES cell, mouse	A-013	5e6	80%		Plasmid (general)	9 µg 100 µl	I/II/2b
	P3	CG-104	5e4	85-95%	58-88%	Plasmid (general)	0.4 µg 20 µl	4D X-Unit

Citations (30)

[An efficient and scalable pipeline for epitope tagging in mammalian stem cells using Cas9 ribonucleoprotein.](#)

Categories: Primary Cells and Media, Transfection

Authors: Dewari PS, Southgate B, McCarten K, Monogarov G, O'Duibhir E, Quinn N, Tyrer A, Leitner MC, Plumb C, Kalantzaki M, Blin C, Finch R, Bressan RB, Morrison G, Jacobi AM, Behlke MA, von Kriegsheim A, Tomlinson S, Krijgsveld J, Pollard SM.

In: eLife (2018) 11;7: 1-29

But to choose which is the best experiment protocol, you must refer to citation papers and test it by yourselves.

